A revision of the *Leucochrysum albicans* (Asteraceae: Gnaphalieae) complex

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Abstract

A morphometric study of the Leucochrysum albicans (DC.) Paul G.Wilson complex (L. albicans, L. graminifolium (Paul G.Wilson) Paul G.Wilson and L molle (DC.) Paul G.Wilson,), concluded that five taxa be recognized from the current six. L. albicans subsp. alpinum (F.Muell.) Paul. G.Wilson is elevated to specific rank as L. alpinum (F.Muell.) R.J.Dennis & N.G.Walsh and L. albicans var. buffaloensis (Paul G.Wilson) Paul G.Wilson is reduced to synonymy under the typical variety. Other members of the complex are maintained at their current ranks.

Keywords: species complex, morphometric analysis, scanning electron microscopy

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Introduction

The Leucochrysum albicans complex comprises a group of closely related taxa that have caused taxonomic confusion in the past; e.g. "Variation between the taxa within Helipterum albicans [now Leucochrysum spp.] appears ... to be continuous ... each taxon intergrades with one or more of the other taxa" (Wilson, 1960, pp. 163, 164). The L. albicans complex is here regarded as consisting of Leucochrysum albicans, L. molle, and L. graminifolium. The two remaining members of the genus, L. fitzgibbonii (F.Muell.) Paul G.Wilson and L. stipitatum (F.Muell.) Paul G.Wilson, are morphologically distinct from the L. albicans complex and both have a Central Australian distribution (Wilson 1992). They were not included in this study.

Leucochrysum molle has a wide distribution including central and eastern South Australia, southern Queensland, western New South Wales, and northern Victoria (Fig. 10). L. molle has linear to broadly obovate, lightly cobwebbed leaves and triangular, ovate to suborbicular yellow involucral bracts. It is an annual whereas other members of the genus are perennials (Wilson 1960). Occurring in sandstone outcrops of the central tablelands in New South Wales, L. graminifolium has a woody stem, usually retaining remnants of previous years' growth. The linear to filiform leaves are light green, the dorsal surface glabrescent while the ventral side remains covered in a cobwebbed mat of hair and the margins are tightly revolute. The involucral bracts are narrow to ovate (Harden 1992). Leucochrysum albicans sens. lat. (sensu Wilson 1992) extends from south eastern Queensland, through eastern New South Wales, the Australian Capital Territory, much of Victoria and into Tasmania.

Leucochrysum albicans sensu Wilson (1992) has two subspecies; albicans, and alpinum, the latter occurring only in the alpine regions of Victoria and New South Wales. Wilson (1992) noted that *L. albicans* subsp. alpinum differs from subsp. albicans by having obovate to oblanceolate, densely lanuginose, flat leaves with slightly incurved margins. The inner involucral bracts are always white and the outer involucral bracts brown to purple, and elliptic to ovate.



Leucochrysum albicans subsp. albicans has linear to oblanceolate, cottony leaves with recurved to tightly revolute margins. It currently includes three varieties: the typical variety, var. buffaloensis (Paul G.Wilson) Paul G.Wilson and var. tricolor (DC.) Paul G.Wilson. The typical variety is widespread throughout the range of the species excluding Tasmania and higher alpine areas. Leucochrysum albicans var. buffaloensis is restricted to Mt Buffalo and immediate environs, Victoria, and var. tricolor has three pockets of distribution, south-eastern New South Wales to north-eastern Victoria, south-western Victoria and scattered sites in Tasmania.

Within *L. albicans* subsp. *albicans*, var. *tricolor* is typically distinguished by the white inner involucral bracts and deep red to purple outer involucral bracts whereas vars *albicans* and *buffaloensis* both have yellow inner involucral bracts and brown outer involucral bracts. Var. *buffaloensis* is distinguished by having acutely tipped, broadly ovate to deltoid involucre bracts whereas var. *albicans* has elliptic to ovate involucre bracts with obtuse apices.

Some of these taxa seem trivially distinguished (Wilson 1960; Short 1999) with some herbarium material being tentatively placed, while subsp. *alpinum* appears to be consistently distinct. In an attempt to resolve the uncertainty apparent within the *L. albicans* complex, a study was undertaken to test the current classification using numerically-based morphometric techniques.

While carrying out this study variation was found in the leaf hairs amongst the different taxa. More detailed study aimed to examine the "stalked glandular hairs" of *L. albicans* and *L. molle* referred to by Short (1999) for any consistent patterns of variation that might support a revised taxonomy. Previous studies of micromorphological characters have found that shape and cell structure of mature carpopodia can be used to delineate taxa (e.g. Sundberg 1985) and these were examined for members of the complex. Scanning electron microscope studies were also undertaken to see if characters of the cypsela pericarp were informative.

Materials and methods Character list and analysis

An initial scoring of 72 specimens using 60 morphological and longevity characters, including those mentioned in

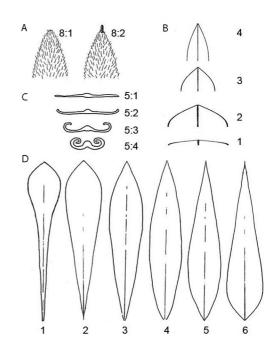


Figure 1: Schematic drawings of (a) leaf tip, showing the two states of character 8; (b) involucral bract lamina tip, showing the range in angles of both characters 12 and 13; (c) leaf margin, showing the range of curvature in character 5; (d) leaf and involucral shape, based on position of widest point along the length of the blade. Characters are numbered as in Table 1.

previous taxonomic treatments (e.g. Wilson 1960, 1992; Harden 1992, Short 1999), was carried out and subjected to analysis using the PATN program package (Belbin 2004). From this analysis, 14 characters were found to be informative and these were used to score 118 herbarium specimens. Previously published references have not separated leaf or involucre bract shape into their component attributes or made reference to other characters found to be informative during this study (Fig. 1; Table 1). These new characters combined with traditional diagnostic characters were used to carry out a morphometric analysis of the members of the *L. albicans* complex.

The 118 specimens examined in the analysis were sourced from the National Herbarium of Victoria (MEL), and the National Herbarium of New South Wales (NSW). Specimens from the herbarium of the Australian Daisy Study Group and collections made in the course of the study by a colleague, K. McDougall, have since

Table 1. Characters for morphometric study.

#	Character	#	Character
1	Life	8	LeafTip
	1. annual		1. tip of leaf covered in hair
	2. perennial		2. dark bald, mucronate tip
2	Leaf shape	9	Length of the lamina of the longest involucral bract (mm)
	1. spathulate		
	2. obovate		
	3. obovate / elliptic		
	4. elliptic		
	5. elliptic / ovate		
	6. ovate		
3	Width of widest part of longest leaf (mm)	10	Involucral bract ratio, length (mm) / width (mm)
4	Leaf ratio	11	Involucral base shape
	leaf length (cm) / leaf width (mm)		1. spathulate
			2. obovate
			3. obovate / elliptic
			4. elliptic
			5. elliptic / ovate
			6. ovate
5	Leaf margin	12	Angle at tip of involucral bract lamina
	1. flat		1. flat to round
	2. curved slightly inwards		2. obtuse
	3. recurved		3. 90°
	4. revolute		4. acute
6	Leaf hair density	13	Angle at base of involucral bract lamina
	1.0 - 10% (glabrescent - sparse)		1. flat to round
	2. 11 - 30% (light cobweb)		2. obtuse
	3. 31 - 60% (cobweb)		3.90°
	4. 61+% (woolly)		4. acute
7	Leaf hair distribution	14	Involucral bract colour
	1. even hair density across dorsal and ventral surfaces		1. yellow
	2. less hair on dorsal surface compared to ventral surface		2. yellow and white
			3. white

been incorporated into MEL's collections. The data were then analysed using PATN. The association matrix utilised the Gower metric. An ordination was produced using Semi-strong Hybrid Multidimensional Scaling (limit= 0.9) with two dimensions and a dendrogram was generated utilising flexible UPGMA (β =-0.1). Evaluation of characters was carried out using Principal Component Correlation (PCC) which generates the value and direction each character has on the points in ordination space (Belbin 2004).

Seedling germination

Plants were grown from seed at the nursery of the Royal Botanic Gardens Melbourne (RBGM). Diagnostic characters seen in the herbarium specimens were compared to seedlings as they were grown in the controlled environment of a glass house.

SEM preparation

Leaf samples were taken from the nursery-grown plants. Plants used were grown for eight weeks. Leaf samples were placed in 70% ethanol upon collection.

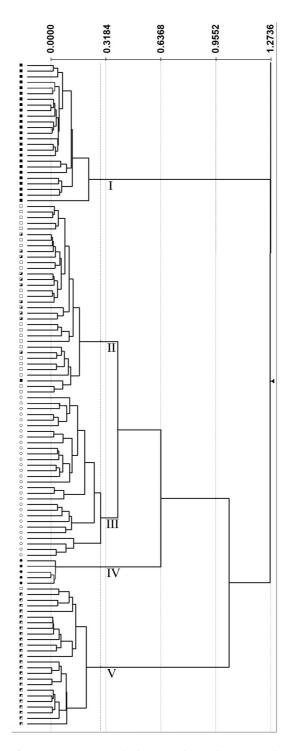


Figure 2: Cluster dendrogram of 118 specimens originally determined as *L. albicans*, *L. molle* and *L. graminifolium*. The 14 characters listed in Table 1 were analysed by Gower metric association with a flexible UPGMA (β=-0.1). At dissimilarity of 0.29 five groups are recognised I-V. Original determinations shown by symbols on left of dendrogram: *L. albicans* subsp. *alpinum* – solid squares; *L. albicans* var. *albicans* – open squares; *L. albicans* var. *buffaloensis* – squares, black in the lower right triangle; *L. albicans* var. *tricolor* – open circles; *L. graminifolium* – solid circles; *L. molle* – squares, black in the top left triangle.

Samples were then treated for 75 min in 90% ethanol then transferred to 100% ethanol and gently agitated for 24 hrs. Samples were then placed in fresh 100% ethanol and agitated for another 24 hrs. Critical Point Drying (CPD) of specimens was carried out the following day following the technique of Boyde and Wood (1969). Leaf tissue taken from the centre of the treated leaves was then placed on SEM stubs exposing both dorsal and ventral surfaces, then coated with gold using a Dynavac 'Xenosput' magnetron sputter coater. A Phillips XL 30 FEG Field Emission Scanning Electron microscope was used to examine the surfaces and capture the images.

From each of the taxa, two to three whole fruits, one with carpopodium exposed were placed on SEM stubs, desiccated, and sputter-coated with gold as described above.

Distribution maps

Distribution data were collated from AD, BRI, CANB, HO, MEL and NSW. Points with a geocode precision of less than 25 km were excluded.

Results

Dendrogram

The composition of the groups largely corresponds with current classification with five groups retrieved above a dissimilarity level of 0.29 (Fig. 2) with Group I comprising *L. albicans* subsp. *alpinum*; Group II, *L. albicans* var. *albicans*; Group III, *L. albicans* var. *tricolor*; Group IV, *L. graminifolium*; and Group V, *L. molle*. PATN could not distinguish *L. albicans* var. *buffaloensis* from var. *albicans*. The dendrogram pairs those specimens most similar to each other first and then progressively unites the groups. The less similar the specimens or groups are the later they will fuse. The first fusion is between Groups II and III, and then Groups IV, V and I successively join the cluster comprised of the other specimens.

Two specimens are apparently misplaced (based on the original determinations). One of these (*Adair s,n.*) originally held at MEL under *L. albicans* subsp. *alpinum* but classified by PATN with Group II (subsp. *albicans*) is a curious specimen, with the white and mauve involucral bracts of subsp. *alpinum* but with leaves narrower than normal and with a glabrous callous tip

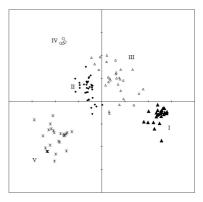


Figure 3: Ordination of 118 specimens originally determined as *L. albicans, L. molle* and *L. graminifolium,* using Semistrong Hybrid Multidimensional Scaling and a Gower metric assocation matrix for the 14 characters in Table 1.

Groups as in Fig. 2.

more characteristic of subsp. *albicans*. It may represent a hybrid between the two taxa, both being in the general area. If so, it is the only hybrid we encountered during the study. The apparently misplaced specimen in Group V (*L. molle*) was simply a misdetermination and this has now been rectified (*Short 3618*).

Ordination

In the ordination the stress was 0.0809, indicating a good fit between the two dimensional representation and the original data matrix. In the ordination space, more similar individuals are placed closer together. The same five groups as identified in the dendrogram (Fig. 2) remain resolved in the ordination (Fig. 3). Groups I (L. albicans subsp. alpinum), IV (L. graminifolium) and V (L. molle) are all well separated, while Groups II and III (L. albicans vars tricolor and albicans respectively) are closely contiguous but discrete. However, when character 14 (bract colour) was excluded from the analysis, groups III and II merged (not illustrated here). This indicates the closeness of these taxa and supports the retention of the bract colour that has traditionally been used to distinguish these two taxa. The composition of the groups is consistent with previously described taxa except in relation to var. buffaloensis. As with the dendrogram, there was no evidence of a discrete group for specimens that had been determined as var. buffaloensis and Group III

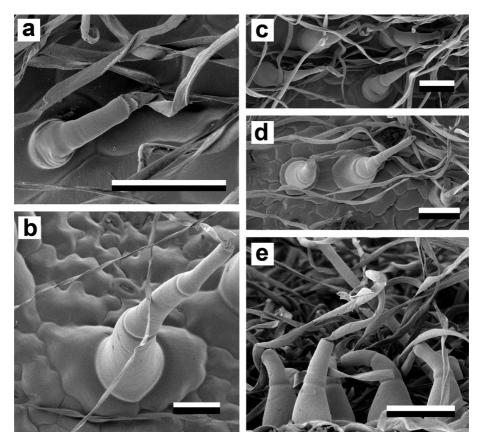


Figure 4: SEM images of the leaf hairs of the dorsal surface of leaves from (a) *L. albicans* subsp. *albicans* var. *tricolor*; (b) *L. molle*; (c) *L. albicans* subsp. *alpinum*; (d) *L. albicans* subsp. *albicans* var. *albicans*; (e) *L. albicans* subsp. *albicans* var. *buffaloensis*. The scale bars are 50 µm.

was a mixture of specimens determined as either var. *albicans* or *buffaloensis*.

Seedling characters

The early-adult leaves of the nursery-raised seedlings generally maintain the diagnostic features apparent on herbarium specimens. However the margins of leaves of all taxa were less strongly recurved than those of herbarium specimens, possibly a consequence of the more humid growing conditions in the nursery.

Leucochrysum molle had leaves with strikingly less dense indumentum, broader than those of var. albicans and much broader those of var. tricolor. The margins of the leaves of L. molle were also less recurved than those of L. albicans vars tricolor and albicans. Leaves of subsp. alpinum were densely covered in hair, had a distinctly spathulate shape, little to no recurving of the margin and

lacked an obvious glabrous mucronate tip. All the other taxa grown have glabrous mucronate tips. There was little difference between vars *buffaloensis* and *albicans*, the former tending to have slightly broader leaves.

Eglandular leaf hairs

The leaf hairs of the taxa examined consist of inflated epidermal cells subtending one to five (usually three) stout basal cells above which is a filamentous ribbon-like extension composed of a number of elongated collapsed cells (Fig. 4). These hairs are found on all taxa examined. They belong to 'hair type B' sensu Drury & Watson (1966) and are common amongst members of the Asteraceae (Bessey, 1889; Bremer, 1994). Leucochrysum molle (Fig. 5b) has the largest hair basal cells of any of the taxa studied. They are at least three times larger than *L. albicans* var. tricolor (Fig. 5a). The

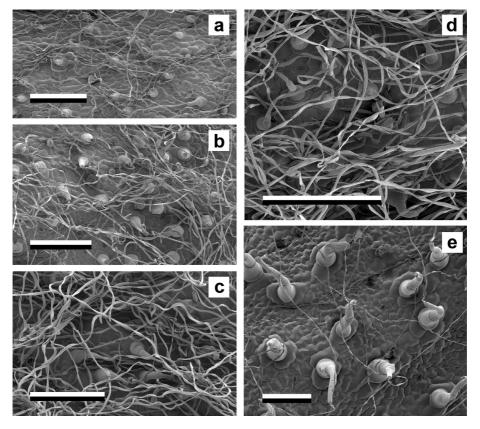


Figure 5: SEM images of the leaf hairs of the dorsal surface of leaves from (a) *L. albicans* subsp. *albicans* var. *albicans*; (b); *L. albicans* subsp. *albicans* var. *tricolor* (c) *L. albicans* subsp. *alpinum*; (d) *L. albicans* subsp. *albicans* var. *buffaloensis*; (e) *L. molle*. The scale bars are 200 μm.

size of the hair basal cells is proportional to the size of the epidermal cells in all the taxa.

The density of hairs differs between the taxa, ranging from the sparsely covered *L. molle* (Fig. 5e) to the densely lanuginose surface of *L. albicans* subsp. *alpinum* (Fig. 5c). The hairs of vars *albicans* (Fig. 6a), *tricolor* (Fig. 6b) and *buffaloensis* (Fig. 6d) are all similarly cottony in contrast. The hairs of the ventral surfaces have longer and thinner basal cells (Fig. 6) than those of the dorsal surfaces (Fig. 4). This difference is consistent across all taxa. The collapsed cells forming the tapering part of subsp. *alpinum* are more twisted than those of the other taxa (Fig. 5).

Glandular leaf hairs

The glandular hairs consist of four rows of paired cells (Fig. 7). The apical secretory pair of cells are larger

than the supporting cells. These hairs are present on all the taxa examined. The general morphology of the glandular hairs is typical of many members of the Asteraceae (Hess 1938). Their distribution is sparse but consistent across the surface of the leaves (Fig. 6)

Pericarp surface

Although there is significant variation in the surface texture of the cypselas examined, both light and scanning electron microscopy has shown these to be of little taxonomic value within the complex. Surface textures examined ranged from smooth to distinctly colliculate, even within a taxon. A selection of SEM images indicating the range of surface textures of the *L. albicans* complex are presented in Figure 8, however, as stated, despite the substantial variation shown here, variability within taxa was too great for this character to

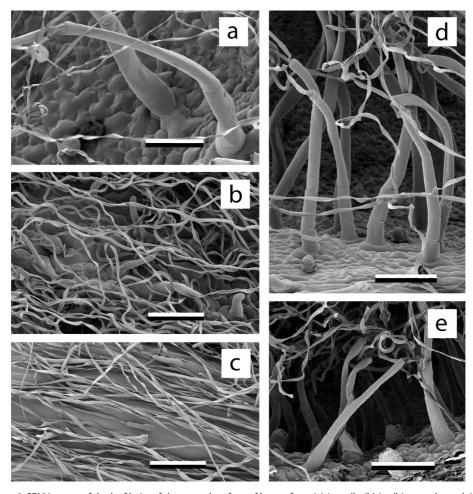


Figure 6: SEM images of the leaf hairs of the ventral surface of leaves from (a) *L. molle*; (b) *L. albicans* subsp. *alpinum*; (c) *L. albicans* subsp. *albicans* var. *tricolor*; (d) *L. albicans* subsp. *albicans* var. *albicans*; (e) *L. albicans* subsp. *albicans* var. *buffaloensis*. The scale bars are 100 µm.

be regarded as useful. The 2-celled myxogenic papillae regarded by Wilson (1992) as being diagnostic of *L. graminifolium* were not observed during this study.

The carpopodium

Scanning electron micrographs are presented of the carpopodia of members of the *L. albicans* complex. There is a distinct boundary between the carpopodium cells and those cells that make up the rest of the pericarp whether the pericarp has a papillose or flat surface (Fig. 9). Despite some variation in outline of the carpopodium (due to the position of the cypsela within the capitulum (Sundberg 1985)) the carpopodia of the different taxa exhibit basically the same morphology.

The carpopodial cells are rectangular to oblong, and between 20 and 60 μ m long.

Taxonomy

Morphological analysis, nursery trials and field examination support the reduction in members of the *L. albicans* complex from six to five, reducing *L. albicans* var. *buffaloensis* to synonymy under the typical variety. The recognition of var. *tricolor* is supported, although on the single criterion of the possession of white rather than yellow capitular bracts. The rank of variety is regarded as appropriate. *Leucochrysum albicans* subsp. *alpinum* is retrieved as a strongly supported group in the analysis. Its distinctiveness in the complex is shown

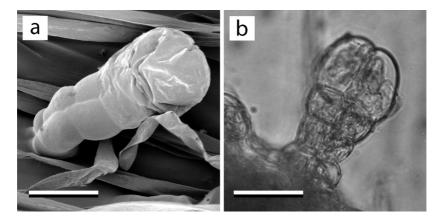


Figure 7: Images of the glandular trichomes on the (a) dorsal surface of a leaf from *L. albicans* subsp. *albicans* var. *buffaloensis* viewed through a light microscope; (b) ventral surface of a leaf from *L. albicans* subsp. *albicans* var. *tricolor* viewed using SEM.

Scale bars are 20 µm.

in both the dendrogram and ordination to be as great or greater than either *L. molle* or *L. graminifolium* from 'core' *L. albicans* and it is proposed that it be recognised similarly at species rank.

The following key serves to distinguish the members of the complex.

Taxonomy and ecology

For full synonymy of the following names, see Wilson (1992).

Leucochrysum albicans (A.Cunn.) Paul G.Wilson, Nuytsia 8:442 (1992)

Helichrysum albicans A.Cunn. in Field, Geog. Mem. New South Wales 359 (1825). Type: Forest Land, Cox's River, 9 Oct. 1822, A. Cunningham 71 (lecto: K; isolecto MEL2068831) fide Wilson 1960.

Leucochrysum albicans var. albicans

Leucochrysum albicans var. buffaloensis (Paul G.Wilson) Paul G.Wilson, Nuytstia 8:443 (1992); Helipterum albicans var. buffaloensis Paul G.Wilson, Trans. Roy. Soc. South Australia 83:170 (1960). Type: Mt Buffalo, Victoria,

Key to taxa

1 Leaves spathulate to broadly obovate, densely woolly; margin flat; apex lacking an obvious mucro or callus tip. Inner involucral bracts lanceolate to ovate, white, outer involucral bracts purplish to brown (especially apparent in early stages of capitulum development)L. alpinum 1: Leaves obovate to filiform, cobwebbed to cottony or glabrescent; margin gently recurved to tightly revolute, apex with short glabrous mucro or callus tip. Inner involucral bracts suborbicular to broadly ovate or ovate, lanceolate to elliptic, white or yellow 2 Annual herb; leaves lightly cobwebbed, obovate to oblancolate. Inner involucral bracts suborbicular to broadly ovate, rounded or truncate at base of lamina, yellow 2: Perennial (although sometimes short-lived); leaves cottony or glabrescent, obovate to oblanceolate 3: Leaves obovate to oblanceolate or linear, cottony; margin recurved to revolute. Inner involucral bracts ovate to lanceolate, white or yellow

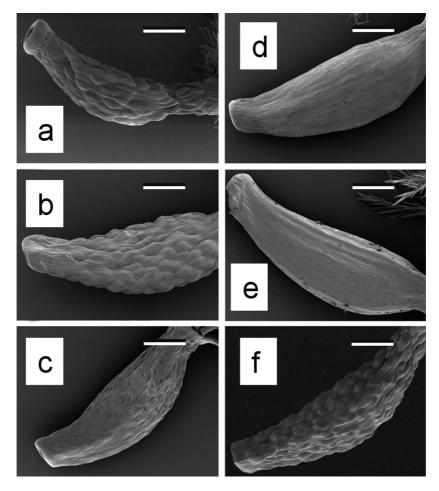


Figure 8: SEM images of cypsela pericarp detail of (a) *L. albicans* subsp. *albicans* var. *tricolor;* (b) *L. molle;* (c) *L. graminifolium*: (d) *L. albicans* subsp. *albicans* var. *buffaloensis*: (e) *L. albicans* subsp. *alpinum*; (f) *L. albicans* subsp. *albicans* var. *albicans*.

Scale bars are 500 μm.

10 Nov. 1955, E. Gauba (holo: GAU) fide Wilson 1992.

Typically associated with dry open forests on shallow, rocky soils derived from a range of parent materials, but frequently sedimentary (mudstones, conglomerates etc.), but also occurring in subalpine heaths and, rarely, lowland grasslands or woodlands. Altitude range c. 100–1800 m. Qld, NSW, ACT, Vic. (Fig. 10a)

Representative specimens (specimens marked with asterisk used for SEM studies): **QUEENSLAND.** Ruby Ck near Stanthorpe, 3.x.1992, *E.Salkin s.n.* (MEL2339313). **NEW SOUTH WALES.** Tenterfield, *C. Stuart s.n.*, *s.d.* (MEL2161748); Near Tenterfield, xi.1874, *C. Stuart s.n.* (MEL2161778); 25 km S of Tenterfield, 29.ix.1987, *E. Salkin s.n.* (MEL2339315); Moonan Brook, 1883, *S.H. Carter, s.n.* (MEL2161764); Timbarra, *C.*

Stuart 136 (MEL); Cavan, near Yass, J.S. Calvert, s.n., s.d. (MEL); New England, collector unknown 30 (MEL); Hastings river, H. Beckler s.n., s.d. (MEL); New England, 1886, R. Collie s.n. (MEL2161790); VICTORIA. Wonga Park, Warrandyte State Park, D.J. van Bockel 88 (MEL); Mt Hedrick, D.E. Albrecht 1952 (MEL292007); Wabonga Plateau State Park, A.D.J. Piesse 393 (MEL685270); Hume Freeway, between Longwood and Euroa, R. Thomas 77 (MEL717321); Mansfield-Longwood Rd, Euroa district, R. Thomas 74 (MEL717324); The Bluff, D.E. Albrecht 1555 (MEL1537304); Old Hume Highway, Winton, J. Strudwick 676 (MEL); 5 km ENE of Chiltern, G. Johnson 25 (MEL1589399); 9.7 km W of Whitfield, H.M. Jolley 17 (MEL2036568, S); Mount Buffalo, Bald Hill, 20.ii.1963, J.H. Willis s.n. (MEL502500); Wall of China, Mt Buffalo, R. Melville 2602 (K, MEL526618); Path to Reeds Lookout, Mt Buffalo, M.A. Todd 250 & 329 (MEL559884, 562279);

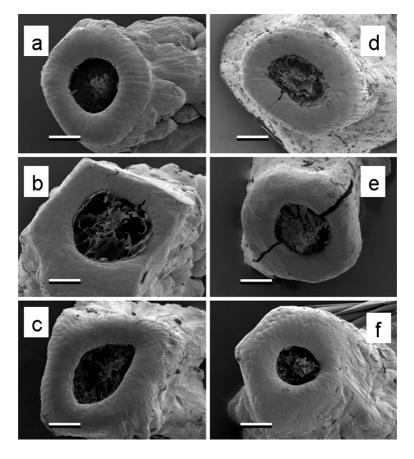


Figure 9: SEM images of carpopodia of (a) *L. albicans* subsp. *albicans* var. *tricolor*; (b) *L. albicans* subsp. *albicans* var. *albicans*; (c) *L. albicans* subsp. *alpinum*; (d) *L. albicans* subsp. *albicans* var. *buffaloensis*; (e) *L graminifolium*; (f) *L. molle*.

Scale bars are 100 μm.

Mt Buffalo NP, *P.S. Short 1403* (MEL601904); Mt Buffalo, *S.T. Blake 7344* (BRI, MEL674708); Mt Buffalo, Chalet, *R. Thomas 76* (MEL717322); Mt Buffalo, *C. & D. Woolcock 1689* (MEL1524031); Mt Buffalo, iii.1931, *P.R.H. St John s.n.* (MEL2162059); Mt Buffalo, iii.1910, *W.R.A. Baker s.n.* (MEL2162060); Mt Buffalo NP, c. 200 m NE from Manfield Lookout, *N.G. Walsh 5977* (MEL); Mt Howitt, 6.ii.1987, *E. Salkin s.n.* (MEL); Eildon-Jamieson Rd, 15 km from Jamieson, 7.xi.1987, *E. Salkin s.n.* (MEL); Old Hume Hwy, Winton, *J. Strudwick 676* (MEL)*.

Leucochrysum albicans var. tricolor (DC.) Paul G.Wilson, Nuytsia 8:443 (1992)

Helipterum incanum var. tricolor DC., Prodr. 6:215 (1838). Type: Van Diemen [Tasmania], R.C. Gunn 108 (holo: G-DC.), fide Wilson 1992.

A plant principally of grasslands and grassy woodlands on relatively fertile soils, often clays or

clay-loams derived from basalt or dolerite, or at higher altitudes, from sedimentary parent material. Altitude range c. 100–900 m a.s.l. NSW, ACT, Vic., Tas. (Fig. 10b)

Representative specimens (specimens marked with asterisk used for SEM studies): NEW SOUTH WALES. 2 km NW of Mt Majura, I.R. Telford 9890 (AD, CANB, MEL, PERTH); S of Tarago, 4.xi.1984, C.E. & D.T. Woolcock s.n. (MEL); Between Blakney Ck and Bevendale, E.M. Canning 6384 (CANB, MEL, NSW, PERTH, S); Hill End, sources of Lachlan River, J. Lauterer 28 (MEL); Wellington, Dubbo, Tomingley, collector unknown 48, s.d. (MEL); Braidwood district, W. Baeuerlen 390 (MEL); Bombay Rd c. 500 m E of Shoalhaven R, K.L. McDougall 1204 (MEL); Kings Hwy, c. 1 km from Captains Flat turnoff, K.L. McDougall 1210 (MEL); Oallen Ford Rd, Windellama, K.L. McDougall 1213 (MEL); Snowy Mtn Hwy, 6 km E of Kosciuszko NP Boundary, K.L. McDougall 1220 (MEL). VICTORIA. Inverleigh Common, E.G. Errey 4861 (MEL); Narrapumelap, A.C. Beauglehole 61646 (MEL); Skipton,



Figure 10. Distribution maps using geocode data records from MEL, AD, CANB and HO for the species (a) *L. albicans* var. *albicans* var. *tricolor*.

W. Whan s.n., s.d. (MEL). **TASMANIA.** Circular Head Bluff, J. Milligan 551 (MEL); ca 6 km SE of Ross, M.J. Brown 45 (HO, MEL); Middlesex Plains, R.C. Gunn 513 (MEL); Circular Head Bluff, J. Milligan 551 (MEL); Circular Head, E. Oakden 115 (MEL); 'van Diemens', R.C. Gunn 155 (MEL); 'Tasmania', G.F. Story 32 (MEL); Rokewood, N.G. Walsh 6619 (MEL)*; Mt Buffalo NP, 0.3 km from McKinnons Corner, J.A. Jeanes 1653 (CANB, K, MEL, NSW)*.

Leucochrysum alpinum (F. Muell.) R.J.Dennis & N.G.Walsh comb. nov.

Helipterum incanum var. alpinum F.Muell., Rep. Pl. Babbage's Exped. 14 (1859); Leucochrysum albicans subsp. alpinum (F.Muell.) Paul G.Wilson, Nuytsia 8:443 (1992). Type: Summit of the Australian Alps, F. Mueller (holo: MEL; iso: K, NSW).

Confined to treeless alpine and high subalpine heathlands and grasslands of the Australian Alps, usually on shallow soils derived from basalt, granite or sedimentary parent material. Altitude range c. 1500–2100 m a.s.l. NSW, Vic. (Fig. 10c)

Representative specimens (specimens marked with asterisk used for SEM studies): NEW SOUTH WALES. 20 km S of Picadilly Circus, W. Bishop 580 (MEL292001, NSW, PRC); Between Seamans Hut and Mt Kosciuszko, G. Stewart 751 (CANB, MEL1582200, PERTH); Scabby Range, c. 2 km SSW of Mt Kelly, F.E. Davies 1712 (CANB, HO, MEL1662341, NSW, PERTH); Seamans Hut, N.G. Walsh 3397 (MEL2013660) Kosciuszko National Park, above Club Lake, P.S. Short 4004 (MEL2014698); 4 km SW from Charlottes Pass, M. Ito 96041 (MEL2030392, NSW); Summit of Mt Clarke, xii.1903, A.J. Tadgell s.n. (MEL2162240); Mt Kosciuszko, R. Helms 121403 (MEL2162242, NSW); Snowy Mountains, W. Baeuerlen 65 (MEL2162244); Mt Kosciuszko, R. von Lendenfield 12 (MEL2162246); Mt Kosciuszko, J. Stirling 23 (MEL2162247); Munyang Mountains, i.1874, F. Mueller s.n. (MEL2162249); Happy Jacks Rd, 2 km from Snowy Mtn Hwy, 25.i.1987, E. Salkin s.n. (MEL2339316). VICTORIA.: Mt Nelse summit, 11.ii.1977, J.H. Willis s.n. (MEL527153); Eskdale Spur, Mt Bogong, L.A. Craven 2127 (CANB, MEL537943); Mt Nelse, 8.i.1954, C. Skewes s.n. (MEL599501); 1.2 km NE of Spion Kopje, 6.ii.1980, R.J. Adair s.n. (MEL678310); Watchbed Ck, A.C. Beauglehole 15635 (MEL1505269); Mount Fainter, A.C.

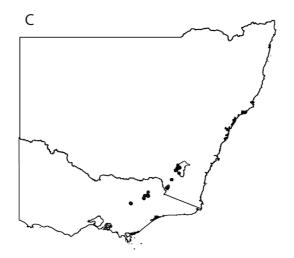


Figure 10 continued. Distribution maps using geocode data records from MEL, AD, CANB and HO for the species (c) *L. alpinum*; (d) *L. molle*; (e) *L. graminifolium*.

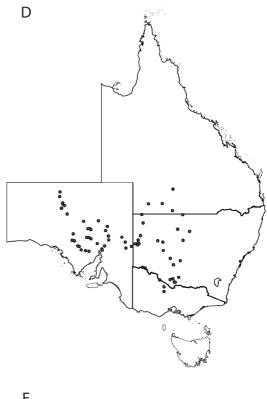
Beauglehole 22494 (MEL1505274); Bogong High Plains, i.1928, A.J. Tadgell s.n. (MEL2162252); Mt Hotham, i.1888, C. Walter s.n. (MEL2162253); Mt Hotham, A.J. Tadgell 75 (MEL2162255); Summit, Mt Nelse North, 25.i.1997, J.Greig s.n. (MEL2339317); Bogong High Plains, c. 100 m SSW of Mt Nelse summit, J.A. Jeanes 1640 (CANB, K, MEL2296401, NSW)*.

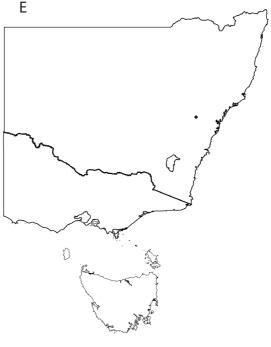
Leucochrysum molle (A.Cunn. ex DC.) Paul G.Wilson, Nuytsia 8:445 (1992).

Helichrysum molle A.Cunn. ex DC., Prodr. 6:194 (1838). Type: Molle's Plains, Lachlan River, New South Wales, July 1817, A. Cunningham (lecto: G-DC) fide Wilson (1960).

A species of semi-arid grasslands, open shrublands or woodlands, commonly with *Acacia* spp. or *Atriplex* spp. as emergents. Soils vary from sands, gibber to fertile clay-loams. Altitude range c. 100–200 m a.s.l. SA, Qld, NSW, Vic. (Fig. 10d)

Representative specimens (specimens marked with asterisk used for SEM studies): SOUTH AUSTRALIA. 3 km S of Pimba, K. Watanabe 326 (AD, MEL2027398, TI); Blinman, Rumball 1402 (MEL2160709). QUEENSLAND. 5 km SW of Eromanga, P.S. Short 3618 (MEL220237); 58.2 km from Quilpie toward Charleville, E.M. Canning 6244 (BRI, CANB, MEL714977, PERTH, S, US); E of Thargomindah, 1885, G.L. Spencer s.n. (MEL2160712). NEW SOUTH WALES. Steam Plains, c. 42 km NW of Jerilderie, T. James 371 (MEL291999, NSW); Sturt NP, S of Olive Downs, W. Greuter 18506 (MEL1543002); 2 km W of Cobar, P.S. Short 3077 (MEL1556462); Tibooburra-Noccundra,





195.8 km from Tibooburra, E. Salkin 762 (MEL2084703); Southern Barrier Range, 1893, H. Deane s.n. (MEL2160681); Warrego River, E. Betche 51 (MEL2160683); Evelyn Ck, N of Barrier Range, 1887, A. King s.n. (MEL2160685); Between the Bogan and the Darling, 1877, W.L. Morton s.n. (MEL2160686); Darling River, 14.x.1860, H. Beckler s.n. (MEL2160689); Between the Lachland and Darling river, x.1885, J. Buckner s.n. (MEL2160691); Lachlan River, G. Tucker 26 (MEL2160692); Silverton, xi.1884, Harris, s.n. (MEL2160693); Lower Edwards River, G.A. Mein s.n., s.d. (MEL2160695); Wilcannia, 1885, M.B. Kennedy s.n. (MEL2160698); Byrock, E. Betche 7 (MEL2160699); Mossgiel, 1885, J. Brueckner s.n. (MEL2160701); Barrier Range, 1889, J.F. Irvine s.n. (MEL2160702); Booligal, ix.1887, A. Bell s.n. (MEL2160706); Yandarlo via Wilcannia, M.B. Kennedy 27 (MEL2160708); Deniliquin, 21,ix.1968, F.J.C. Rogers s.n. (MEL2231783). VICTORIA. Kerang Mitiamo Rd, 22.xi.2005, D. Marshall s.n. (MEL2293160)*.

Leucochrysum graminifolium (Paul G.Wilson) Paul G.Wilson, Nuytsia 8:444 (1992).

Helipterum albicans var. graminifolium Paul G.Wilson, Trans. Roy. Soc. South Australia 83:171 (1960). Type: Clarence-Wolgan Rd, New South Wales, 26 Feb. 1939, W.F. Blakely, J. & W.J. Buckingham 3306 (holo: NSW; iso: AD. MEL).

A localised endemic growing on sandstone-ironstone outcrops (known locally as 'pagodas') in the Newnes-Capertee Valley area. Altitude range c. 800–1000 m a.s.l. NSW only. (Fig. 10e)

Representative specimens: NEW SOUTH WALES. Clarence-Wolgan Rd, 31.xii.1939, W.F. Blakely, J. & W.J. Buckingham s.n., (MEL, NSW); Wolgan Gap, 12.iv.1953, L.A.S. Johnson s.n. (NSW); Newnes Plateau, J. Porter et al. 20119 (MEL291446, NSW); Glowworm Tunnel Rd, Wollemi National Park, M. Kennedy et al. 59 (NSW).

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References

- Belbin, L. (2004) *PATN, pattern analysis package*. Blatant Fabrications Pty Ltd.: Tasmania.
- Bessey, C.E. (1889) Botany: for High Schools and Colleges; from the American Science Series Advance course. Henery Holt and Co.: New York.
- Boyde, A. and Wood, C. (1969) Preparation of animal tissue for surface scanning electron microscopy, *J. Microsc.* **90**, 221–249.
- Bremer, K. (1994) *Asteraceae; cladistics and classification,* Timber Press: Portland, Oregon. pp: 339–349.
- Drury, D.G. and Watson, L. (1966) Taxonomic implications of a comparative anatomical study of Inuloideae-Compositae, Amer. J. Bot., 53, 828–833.
- Harden, G.J. (1992) Flora of New South Wales, UNSW Press Ltd, pp: 227–228.
- Hess, R. (1938) Vergleichende Unteruschungen über die Zwillinghaare der Compositen, *Bot. Jarb. Syst.* **68**, 435–496.
- Lindley, J. (1836) A natural system of botany, edn 2. Longman, London.
- Short, P.S. (1999) Leucochrysum. In Walsh, N.G. & Entwisle, T.J. (eds). Flora of Victoria vol. 4. Inkata Press, Melbourne. pp: 789–792.
- Sundberg, S. (1985) Micromorphological characters as generic markers in the Asteraceae, *Taxon*, **34**, 31–37.
- Wilson, P.G (1960) A consideration of the species previously included within *Helipterum albicans* (A.Cunn.) DC., *Translations of The Royal Society of South Australia*. **82**, 163–177.
- Wilson, P.G. (1989) A revision of the genus *Hyalosperma* (Asteraceae: Inuleae: Gnaphaliinae), *Nuytsia* **7**, 75–101.
- Wilson, P.G. (1992) The classification of Australian species currently included in *Helipterum* (Asteraceae: Gnaphalieae): Part 2 *Leucochrysum*, *Nuytsia* **8**, 439–446.