



## AUSTRALIA'S FUNGI MAPPING SCHEME

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### THANKS FOR A GREAT EVENT: FUNGIMAP VI 2011

*Jarred Pedro*

The Fungimap VI Conference was recently held over five days at the WA College of Agriculture in Denmark, Western Australia.

The Conference kicked off on Thursday 14th July with delegates arriving from all over Australia and indeed the world - bringing together old friends and colleagues and providing the opportunity to make new ones through the sharing of knowledge, exchange of stories and the united expectation of what was to come over the next few days.

The conference was full of interesting talks, educational workshops and fun forays.

Throughout the conference there was a range of social activities including a celebratory dinner, with some great food and great company.

The Denmark Agricultural College proved to be a great venue with 64 of the 95 people participating in the full conference staying on site at the College. This provided the opportunity for further networking and sharing of knowledge in a fun and relaxed atmosphere.

The conference was a great success, however this was no wonder considering the amount of time and effort generously contributed by the Fungimap conference committee and volunteers.

The conference created a valuable opportunity for fungi enthusiasts to get together, share their knowledge, enjoy each other's company and generally have a great time.

See Page 8 for Jarred's detailed overview of the Fungimap VI Conference activities.

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## FROM THE EDITOR

Hello! This is the first *Fungimap Newsletter* that I've had the privilege of working on. I am lucky to become involved at a time when so much is happening. The reports back from Fungimap VI have been very positive – I have even heard whispers of 'best-ever' on the grapevine. This edition comprehensively covers the gathering – with reports and photos of the academic proceedings, social activities and accompanying forays provided by members.

I am happy to be taking the reins from your newsletter editor of the last 6 years, Pam Catcheside, who has generously shown me the ropes this year. Pam's dedicated service, fungal expertise and passion has produced so many wonderful newsletters. Her time as newsletter editor is just one example of the tireless efforts that led to Pam's award last year as an *Unsung Hero for South Australian Science* by the Government of SA. I am sure that future newsletters will continue to feature Pam's achievements and valued contributions to the Fungimap community.

Thank you to all contributors whose efforts make up this issue. It's not too early to start thinking about sending articles, news items, and fungi images for our bumper December issue. Please remember that images should be least at 300 dpi and approx. 1Mb in size, and always accompanied by a copyright release. This is as simple as popping a statement (see the box on Page 5) into the email when you submit your photos.

**Please note: the deadline for the next issue is Friday, 2nd Dec 2011.**

Happy reading!

**Christina Hall.**

## FUNGAL NEWS

For a list of **Fungi events** and activities around Australia, see the [calendar of fungi events](#) on the Fungimap website, where you can also find contact details of the various [regional fungi groups](#).

### News from the Queensland Mycological Society

*Sapphire McMullan-Fisher*

QMS has had a full year with a bi-monthly meeting schedule and nearly monthly forays until August. We now slow down a bit for the 'dry season' continuing our bi-monthly meetings but only having a workshop before the next wet fungal season!

The start of 2011 was not great with the La Nina driven weather over summer badly affecting Queensland and some of our members' families and homes were damaged by the floods and cyclones. Luckily no lives were lost.

Also early 2011 brought the news that Myrtle Rust (*Uredo rangellii*) had been found in both Queensland and New South Wales. Unfortunately it seems that this disease, although detected in 2010, may have been present in Australia for at least two years. This disease probably came from Central and Southern America and is a group of fungi known as the 'guava rust complex.' This disease affects members of the *Myrtaceae* family of plants, which includes many Australian native species from genera including *Acmena*, *Austromyrtus*, *Backhousia*, *Callistemon*, *Eucalyptus*, *Gossia*, *Leptospermum*, *Melaleuca*, *Rhodamnia*, *Syncarpia*, and *Syzygium*.

Although this disease will not be able to be stopped, the spread can be slowed. So all QMS members have been encouraged to practice good hygiene when moving between sites. Members arrive at sites with clean equipment (shoes, tripods, spades, cars etc.) and clothes (including hats) for forays and other activities. Particular care was taken not to spread Myrtle rust by members who went to WA for the Fungimap Conference in July, as Myrtle rust has not been found in WA yet.

The two highlights from this year's talks so far have been the talk from Dr Ross McKenzie on 'Mycotoxicosis Oz: A survey of poisonous Fungi in Australia' (8 February 2011). And the QMS AGM in April when Katrina Syme (Botanical Artist and Mycologist) spoke about 'Fascination with Fungi'. Katie took us on a delightful ramble about her discovery, and developing interest in fungi, which has taken her all over the world. By the time of publication, Queensland Herbarium mycologist Nigel Fechner will have spoken (11 Oct 2011) about his and Roy Hallings 'Fraser Fungi Project.'

### Central Coast news

*Pam O'Sullivan*

Do you enjoy taking photos of fungi and not really worry or even think that these images may have any long term scientific or historic value? After all you may not be collecting specimens to be fully documented and lodged in a herbarium. Have you ever wondered if documenting the fungal species in your area is just a bit like stamp collecting and more about self-indulgence than of any real scientific value? Well think again!

Recently, on the Central Coast of NSW we have heard that one of the areas we foray in each year, Hargraves Beach, is being earmarked by the local Wyong Council for an Eco Village of 50 units! There are all of the environmental issues that arise with such a project in a delicate hind dune area, coupled with a potential rise in the sea level. However, for us there is the loss of biodiversity, in particular the fungi! It is an area that the dunecare group has been doing weed removal, restoration and documentation of the biota since the 1990's. Our group has assisted in documenting the fungi at the site for the last three years.

It is always an exciting site to visit as we always find fungal species there that we don't find at any of our other foraging sites in the region. Unusual species, such as the Ascomycota *Hypocrea gelatinosa* - an incredible little translucent jelly-like blob with dark dots of spores masses within the fungus. These tiny fruiting bodies were found scattered across a well rotted log. We think this is the only recording for NSW! The infrequently recorded *Myriostoma coliforme*, known in England as the Pepper Pot, which, unlike most 'earthstars', has 5-10 tiny necks or stems between the outer lobes (exoperidium) that hold the fungus off the ground and the globose spore bearing endoperidium. The endoperidium has multiple pores (like a pepper pot) through which the spores are released unlike the single pore seen on most *Gastrum* species. We recorded another puffball, the hard skinned *Mycenastrum corium* only at this site as well.

There are many, many other wonderful species we have recorded and photographed at Hargraves Beach - so regardless of what happens to this little piece of land there will be a small record for future generations of some of the species that were part of this fragile ecosystem. This was a reminder to us, and we wanted to share this with you all: when you least expect it, the humble efforts of getting to know and enjoy the fungi in our areas and recording them, has long term and significant value. So keep enjoying, photographing and recording!

## News from SA

Pam Catcheside

It has been a varied year for the Adelaide Fungal Studies Group. Apart from the traditional forays and meetings, we have held a workshop in the south-east of South Australia, taken the Bellevue Heights Junior Field Naturalists' group on a foray to Belair National Park and given an information session at the Adelaide Farmers' Market organised by the Royal Institution of Australia.

After our somewhat decadent first foray of the season to Mount Lofty Botanic Garden when we concluded our foray with a lunch in nearby Stirling, we settled into serious fungologising. The Lepiotes were predominant at our late May foray. Collections included the small, all white *Lepiota minya* and *Lepiota haemorrhagica*, its cap and stipe covered with red-brown fibrillose scales, its creamy white gills becoming red-orange when bruised.

Our workshops in early June involved surveys of Conservation Parks and Forestry Reserves near Mount Gambier in the State's south-east. Jelly Babies, *Leotia lubrica*, with its greenish, gelatinous knobs on yellow stipes was abundant. There were some splendid specimens of apricot-coloured waxcap, *Cantharellus cinnabarinus* var. *australiensis*. Lilac fruit bodies of *Humidicutis lewellinae*, their thick, waxy caps splitting and splaying at the margins, and *Humidicutis viridimagentea* (Figure 1) with red-purple caps, yellow gills and multicoloured stipes were amongst the collections; the latter species has green caps when young.



Figure 1. *Humidicutis viridimagentea*  
(Photo: David Catcheside).

Another find was *Tricholomopsis rutilans*, its custard yellow gills and flesh, its cap and stipe ornamented with plum coloured scales giving it the common name of Plums and Custard. On a much smaller scale were two tiny orange disc fungi on dung, *Cheilymenia raripila* on wombat dung and *C. coprinaria* on that of grey kangaroo. The species are, in part, separated on the size and delicacy of the dark hairs on the outer surface of the receptacle. *C. stercorea* has conspicuous hairs that are distinctly forked at their bases, while *C. raripila* has delicate, inconspicuous simple hairs that are not distinctly forked. At first, I was delighted to find tiny fruit bodies of *Pilobolus* amongst the discs of *C. raripila*. However, I was not so delighted to observe that the white 'hairs' around the discs were not hairs but writhing nematodes, and likely parasites within their vertebrate host. The relationship between *Pilobolus* and the nematodes is fascinating: *Pilobolus* comprises a translucent white stem less than 10 mm high; its base is embedded in the dung, the stalk terminates in a balloon-like vesicle, on top of which is a shining, black, flattened sphere, the sporangium (spore case). The stalk is phototropic, bending towards the light; the vesicle is full of liquid and, when the pressure has built up, the sporangium is launched up to 2.5 m from the dung surface. Mucilage surrounding the spore case enables it to stick to plant material where it is more likely to be eaten by passing animals - few animals choose to eat their own dung. The parasitic nematodes make use of the *Pilobolus* by climbing on to it and may subsequently be shot with the sporangium and eaten by grazing animals. On noting the nematodes, I was especially careful to wash my hands and clean all equipment very thoroughly!

In mid-May we were accompanied by a number of eager members of the Bellevue Heights Junior Field Naturalists' Society, their parents and Rona Sakko, the Convener of the group (Figure 2).



Figure 2. Junior Field Naturalists at Belair National Park (Photo: Uni Carnegie).

Our first find of a nest of truffles, *Hysterangium inflatum*, caused some excitement and after that there were frequent calls of 'Come and look at this!' It was a great morning

and we hope to have further forays with the Juniors next year. One of the finds exemplified another story relating to invertebrates. The narcotic effects of the Fly Agaric, *Amanita muscaria* are well-known. In mediaeval times, saucers of milk into which fruit bodies had been crumbled were placed on window sills as fly catchers. The effects of the fungus were demonstrated by aggregations of dead flies on the rings of *A. muscaria* fruit bodies (Figure 3).



**Figure 3. *Amanita muscaria* with flies**  
(Photo: David Catchside).

Finally, an example of biological control: a grasshopper, collected in Penambol Conservation Park near Mount Gambier (Figure 4).



**Figure 4. Parasitised grasshopper**  
(Photo: David Catchside).

#### References

- Kendrick B (1992). *The Fifth Kingdom*. Mycologue Publications, Ontario, Canada.
- Ramsbottom J (1954). *Mushrooms and Toadstools. The New Naturalist Series*, Collins, London.
- Young AM, Syme K (2007). A new green species of *Humidicutis* from Western Australia. *Australasian Mycologist* 26: 71-74.

Do you take amazing photos of fungi like these? Please share them with the Fungimap community by sending them to [ChristinaRHall@yahoo.com.au](mailto:ChristinaRHall@yahoo.com.au) for inclusion in the *Fungimap Newsletter*.

Our preferred option is for a Fungimap record sheet to be submitted along with images. This means that the identification and locality of the fungus can be permanently stored in the Fungimap database. If you submit photos this way, you can choose to accept the copyright statement on the records form. You can download record forms at: <http://www.rbg.vic.gov.au/fungimap/submitting-fungi-records>

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## HOW DO FUNGI FARE IN *Australia's Biodiversity Conservation Strategy 2010-2030?*

Tom May

*Australia's Biodiversity Conservation Strategy 2010-2030* was released by the Commonwealth Department of Sustainability, Environment, Water, Population and Communities in December 2010, at the tail end of the International Year of Biodiversity. It is a significant document that will be the 'guiding framework' for conserving Australia's biodiversity over the next two decades.

Fungimap submitted a response to the draft Strategy in 2010. Our main points in this submission and the eventual treatment in the Strategy are summarised in the following table.

<b>Fungimap Submission</b>	<b>Biodiversity Conservation Strategy</b>
More background information needed on what is included under 'biodiversity', with an overview of the diversity of life, mentioning all biota including fungi, protozoans and bacteria, what they are doing in ecosystems and how they interact. For each major group, the number of known species should be provided, as well as the number yet to be described, and levels of endemism	Very brief treatment of 'biodiversity' with no specifics other than it includes 'plants, animals and micro-organisms'. Mentions that there are as many as 560,000 species in Australia, but no breakdown provided. Endemism levels provided for some groups of plants and animals. The words 'fungi' or 'fungus' do not appear in the body of the report.
In order to effectively conserve biota there is a need for baseline information about what species there are, where they occur, and how many are yet to be described. Therefore, need to create inventory of all known Australian species (with rigorous estimates of unknown species) by 2015, and describe 80% of known species that lack formal names by 2020.	'By 2015, nationally agreed science and knowledge priorities for biodiversity conservation are guiding research activities.' However, there are no specific targets for inventory or description of species, and a lack of detail about the scope of the 'priorities'.
Most fungi are not yet even collected, documented and formally named. Therefore create, test and refine explicit strategies for conserving poorly known groups such as fungi. Such strategies could include use of other biota as surrogates for fungi.	Totally ignored. The words 'surrogate' or 'umbrella' do not appear in the Strategy.
There has been no comprehensive threat assessment for fungi nationally or for individual states. Therefore carry out a comprehensive threat assessments across all known species by 2015 so that we know which species are endangered, and can direct resources appropriately, and track changes in threat status.	No mention at all of threat assessments.
Commendable aim in the draft Strategy of building baseline datasets and setting up nationally representative set of long-term monitoring protocols and sites. However, need to be explicit that fungi (and other lesser known groups) must be included in such datasets and monitoring protocols.	Establishment of 'national long-term biodiversity monitoring and reporting system' is a goal, but no mention that it needs to cover all biota.
There is great potential to harness networks of volunteers to collect distribution and ecological information that is not otherwise available. However, to maximise the input of volunteers and community groups in biodiversity monitoring there must be a willingness to support these groups, both financially and by providing standards for data collection and management.	Volunteers as such are omitted. There is mention of community involvement in several places, but no mention of government support for community groups engaged in biodiversity conservation. 'Enhancing strategic investments and partnerships' focuses on an increase in three areas: (1) markets and other incentives for managing biodiversity and ecosystem services, (2) private expenditure on biodiversity conservation, and (3) public-private partnerships for biodiversity conservation. It is not clear how any of these 'investments' will directly assist volunteers.

The lack of attention to fungi in the Strategy is epitomised by the fact that the only place the word 'fungi' appears is in the caption to an image (p. 65). Although merely labelled 'bracket fungi' the species depicted is clearly the Fungimap target species *Stereum ostrea*.

In the Strategy, targets and actions are arranged under three priority areas: (1) Engaging all Australians; (2) Building ecosystem resilience in a changing climate; and (3) Getting measurable results. Each priority area has three subpriorities, and there are national targets for each by 2015. There are also 26 actions arranged under the three priority areas.

For the 2015 targets, there are various worthy aims in terms of 'national increase of 600,000 km<sup>2</sup> of native habitat managed primarily for biodiversity conservation' and establishing 'four collaborative continental-scale linkages'. However, due to lack of knowledge, fungi are unlikely to explicitly benefit from any of these measures, although individual species and communities of fungi may unwittingly benefit (but without specific monitoring how would we know?).

The most relevant actions for fungi include:

- A14. Identify and protect climate change refugia ...
- A19. Complete (as in finish) assessments at national state and regional levels to identify knowledge needs and gaps and to set research priorities for biodiversity conservation at all levels.
- A20. Develop national biodiversity account in conjunction with broader national environmental accounting and reporting systems.
- A23. Implement ... conservation models that apply a whole-of-ecosystem approach ...
- A24. Develop ... a nationally representative set of biodiversity indicators and monitoring protocols.

For 2015, the national targets that relate specifically to these goals are simply:

- nationally agreed science and knowledge priorities for biodiversity conservation are guiding research activities;
- all jurisdictions will review relevant legislation, policies and programs to maximise alignment with Australia's Biodiversity Conservation Strategy; and
- establish a national long-term biodiversity monitoring and reporting system.

These targets just seem to put off for another five years actually setting any concrete goals. Our concluding comment in the submission was 'Overall, there is a lack of specifics in the Strategy. There is a distinct risk that general statements of objectives are met sufficiently over the life of the Strategy to tick them off as having been achieved, but only in a general sense that across all governments there is some progress in some areas ... surely a national Strategy should be where some of the specifics of the priorities are set now, such as for targets for the number of species to be described, or the proportion of species that are covered by threat assessments.'

If anything, there is less detail in the final Strategy than in the draft. As far as fungi, the document is disappointing. There is a real danger that fungi will continue to be overlooked due to the lack of integration of them into existing programs. The fact that there are very few fungi formally listed under State and Commonwealth endangered species legislation already retards funding and resources directed at research, management and education about fungi. It will be important to ensure that fungi are included in actions under the Strategy that do offer some prospect of improving the situation - in particular, the establishment of biodiversity monitoring and reporting systems.

Given the lack of specifics about fungi in government biodiversity strategy documents (the situation for most States is similar to the Commonwealth) it is time to start planning for creation and monitoring of a strategy about documentation and conservation of Australian fungi, to be produced by non-government groups. The 1997 Conservation Overview of Australian non-marine lichens, bryophytes, algae and fungi did contain specific recommendations for fungi and the other groups covered. However, there was no follow up of the recommendations in the Overview.

Creation of a Fungi Strategy by non-government groups would allow for a wide-ranging consideration of what really is required to adequately document, understand and conserve Australian fungi. In addition, monitoring of real progress could be carried out, by keeping track of indicators such as numbers of species described, proportion of species with threat status assessments, numbers of staff in conservation agencies with responsibility for fungi, and so on.

Australia's Biodiversity Conservation Strategy 2010-2030 can be downloaded from: <http://www.environment.gov.au/biodiversity/publications/strategy-2010-30/index.html>  
Fungi map's response to the draft Strategy is available at <http://www.rbg.vic.gov.au/fungimap/policy-development>

## FUNGIMAP VI



Conference attendees at the Fungimap VI day of talks (Photo: Bernard Wong).

### The conference

*Jarred Pedro*

The Fungimap VI Conference took place in Denmark, south west of WA at the WA College of Agriculture. This was quite a special Fungimap Conference for me because I had attended the very first conference held here in Denmark ten years ago in 2001 when I was only 9 years old.

The conference commenced on Thursday the 14th of July with delegates converging from far and wide. After a shared dinner there was a most intriguing presentation by Donna Franklin, a PhD candidate and tutor in Cultural History and Theory at ECU. Donna shared her interest and obvious passion in creating art works that explore the use of fungi, plants and animals - including a dress made of fungi.

On Friday the 15th of July 110 people attended the day of talks held in the Denmark Civic Centre. It consisted of 13 presentations by national and international mycologists and fungi enthusiasts covering diverse subjects. Kevin Thiele's 'Making Biodiversity information available to the public' showed us the numerous information options available on the net, and Keith Bradby helped us to understand what really matters in our environment and how important it is to look after it. Then we were transported to the arid lands by Arpad Kalotas with his presentation on the uses of fungi by the indigenous people of the desert regions.

Neale Bougher gave a talk on the fundamental relationships between fungi and the Gilbert's Potoroo, as well as similarly important relationships with Beetles outlining some remarkable research findings. Richard Robinson provided an insight into fungi's relationship with fire and Mark Brundrett thrilled us all with some amazing photographic images that helped to explain the remarkable global significance of beneficial plant-fungus associations.

Every one took a breather for lunch before launching back into the second half of this educational journey. To kick it off the ever-enthusiastic Laurton McGurk gave an entertaining overview of her beloved Amanita research. Brandon Matheny showed us there was more to Inocybaceae than most of us probably realized and Pete Davison got so enthused about his slime moulds that he was hard to stop!

The theme then changed as Kirsten Tullis treated us to a virtual walk through the bush before Teresa Van der Heul also talked us through a photographic journey of their local bushlands. Sapphire McMullan-Fisher presented a summary of the fungi survey she had completed in the Mary Cairncross Scenic Reserve and finally Paul George gave us an update of the most common and the rarest species found over the past 8 years while conducting over 70 forays in 20 locations.

Saturday the 16th and Sunday the 17th of July consisted of forays in surrounding bush land and private property. Each morning every one organized themselves into 10 groups and headed off to find and record fungi - over the two days the 20 groups recorded approximately 240 species.



Foray leader Paul George (Vic) points out the finer details to enthusiastic participants (Photo: Patricia Gurry).



Jarred Pedro (WA) gets up close and personal with fungi (Photo: Patricia Gurry).



Participants on the foray to the Mercer property (Photo: Patricia Gurry).

In the afternoon workshops were held at the WA College of Agriculture, with the workshop leaders coming from diverse backgrounds and providing topics ranging from beginner workshops on basic fungal identification, through to skills such as writing descriptions, taking photos of, and surveying macrofungi. There were also workshops identifying the microscopic features of specific fungi groups.



Patrick Leonard presents his *Russala* & *Lactarius* Identification workshop (Photo: Bernard Wong).

On Monday the 18th July a foray was organized to spend the day at the Tingle Forest and Tree Top Walk, 50 km west of Denmark. This gave everyone the opportunity to survey the fungi in the unique Karri and Tingle forest, a globally renowned biodiversity hot spot.



Tree Top Walk - Virgil Hubregtse, Pam O'Sullivan & Yvonne Maher (with Elaine Davison behind) (Photo: Laurton McGurk).

Socialising during the evenings included a celebratory dinner as well as a Fungimap trivia night. By all accounts, the trivia questions were full of the most interesting information and little-known facts, thanks to Ian Bell.



Trivia Night. L-R at table: Alex Syme, Blanche Higgins, Julie Fielder, Diana Leemon, Karen Clarke and Mark Brundrett. Ian Bell in background (Photo: SJM McMullan-Fisher).

So much hard work goes into the preparation of a Conference, so I would like to thank the committee for their commitment and hard work in putting together such a great experience.

## Foray Report

Sapphire McMullan-Fisher and Katrina Syme

The sixth Fungimap Conference in Denmark proved to be a very successful event. One of the excellent aspects was the fungi forays, held at 20 sites on Saturday and Sunday, which were a great learning opportunity for all. Most of the sites were in reserves, but three local property owners generously made their places available to us. On Monday we were bussed westwards to the Walpole-Nornalup National Park's Tree Top Walk in the Valley of the Giants for a more relaxed foray amid the magnificent Red Tingle trees.

To summarise the results of the macrofungi observed during the conference forays:

- 665 observations were made of macrofungi from 20 morphogroups of fungi from 3 phyla (*Ascomycota*, *Basidiomycota* and *Myxomycota*).
- 109 records were made of 22 Fungimap target species

Of the total 235 taxa there were:

- 130 named species and 19 species equivalents
- 72 taxa recognised at the level of genus
- 14 taxa recognised to morphogroup only (e.g. '*Discomycete* sp. orange')



*Anthracophyllum archeri* was one of the most common Fungimap target seen during Fungimap VI (Photo: SJM McMullan-Fisher).

Only a few species were seen at more than half the sites. These included Fungimap target species *Amanita xanthocephala* (15 sites), *Anthracophyllum archeri* (17) and *Stereum hirsutum* (14).

Other commonly recorded species were *Coltricia cinnamomea* (19), *Hydnum repandum* (17) and *Inocybe* sp. (15). It is interesting that some very common taxa were found across the different vegetation types. There are no obvious species favouring any particular dominant tree (see Table 1 in Fungimap Bulletin 2, where sites are grouped by dominant tree species).



*Hydnoplicata convoluta* (Photo: Katrina Syme).

As for most survey data sets, one hundred and twenty three (or nearly half) of the taxa were only seen at a single site. The most species-rich site had 73 taxa recorded and at the least rich two sites only 37 taxa were recorded. Although morphogroups are not strict scientific categories, they are useful for grouping fungi, as they are based on their shapes. As is typical, the gilled fungi (114 taxa) formed slightly more than half the total observations, which comprised 235 taxa. The rest of the taxa were made up of the other 19 morphogroups. This data set includes a good number of Sequestrate or Truffle-like fungi (13 taxa), found because of the knowledge of this group by some of the leaders and participants. Most data sets are biased towards the groups of fungi about which people know the most so the accumulated experience of conference participants resulted in a very broad data set across the macrofungi.

The collection and distribution of this data in such a timely manner was made possible by the effort of all of the leaders, the recorders and Geoff Lay - who entered the data during the conference.

The full species list as a table of which species were at which sites (Table 1), along with site details (Table 2), is available as Fungimap Bulletin 2 on the Fungimap website:

[http://www.rbg.vic.gov.au/\\_data/assets/pdf\\_file/0006/16098/Fungimap\\_Bulletin\\_2.pdf](http://www.rbg.vic.gov.au/_data/assets/pdf_file/0006/16098/Fungimap_Bulletin_2.pdf)

Although this is observational data (and some named species need verifying), it does demonstrate that in three days a concentration of eyes on the ground together with experienced leaders, many records can be made. The data is being shared with the Western Australian Department of

Environment and Conservation, and anyone else who is interested in the results.

Many of the participants reported that they really enjoyed the conference forays. The sites were in a variety of very good local vegetation, which was new habitat for most of the participants. There was a wide range of fresh fungi to admire, photograph and learn about. As for the whole conference, the camaraderie was wonderful - and last but not least the weather was sunny if a little chilly with Denmark's first frost in years.



**Foray participants Teresa Van Der Heul (N.S.W.) and Yvonne Maher (S.A.) identifying fungi (Photo: Patricia Gurry).**



***Dermocybe erythrocephala* (Photo: Katrina Syme).**



***Dermocybe austroveneta* (Photo: Patricia Gurry).**

***Make sure you head to the Fungimap website to see the exquisite colours of these specimens found on the Fungimap VI forays!***

***Fungimap Newsletters are available for viewing in pdf format with all images in colour at:***

**<http://www.rbg.vic.gov.au/fungimap/fungimap-newsletter>**



***Cortinarius rotundisporus* (Photo: Patricia Gurry).**

**BOOK REVIEW:****Pocket Guide: Mushrooms of South Africa**

Marieka Gryzenhout 2010

ISBN 9781770077560, Struik Nature

Softcover: 144 pages.

\$29.95 + \$3 postage from the Fungimap Bookshop.

*Reviewed by Sapphire McMullan-Fisher*

This pocket size field guide would be a useful tool for any naturalist visiting South Africa. The introductory pages provide a solid introduction to fungi and their roles in ecosystems. It includes some information about the shapes of common fungi, the morphological groups used in the guide and fungi generally. There are also useful name and species indexes at the end of the guide.

Unfortunately, nowhere does the author standardise sizing, so what defines small, medium or large fungi are left to the reader to interpret. Fortunately the use of objects including rulers, trowels, an impressive knife, secateurs etc. as scales in many of the pictures helps the reader get a feel for the size of some fungi, but it would have been more useful if a consistent and universally understandable scale was used.

The fungi are grouped by morphological characteristics, based on a simple series of descriptions of these morphological groups, and is easy to use. The scientist in me is a bit perplexed by the order used for these morphological groupings within the book; similar shaped groups are not necessarily near each other, although the author suggests they are, so perhaps we group fungi differently?

Chanterelles and Toothed fungi, which usually have caps and stems, are not grouped near the mushrooms. No phylogenetic order applied in the guide; the Ascomycetes are scattered amongst a few Basidiomycetes towards the end of the book. My copy of the guide has had the morphological grouping pages corrected, so clearly defining the order of the book has been problematic. For future editions it would be nice to see similar shaped fungi near each other.

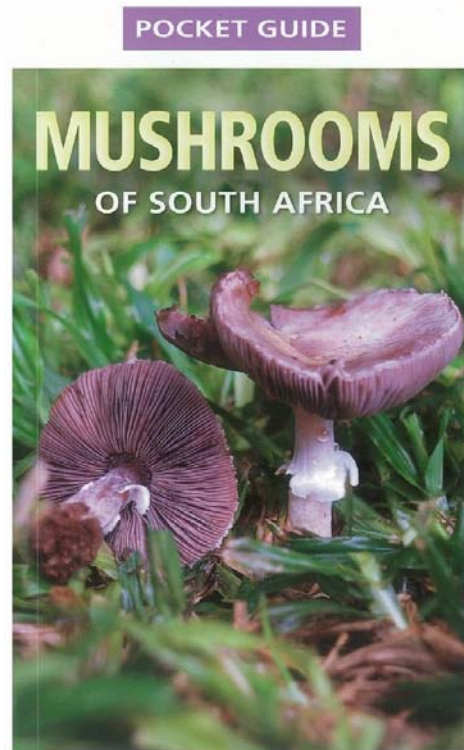
The author is clearly a capable mycologist; the text holds many references to the need to consider microscopic characteristics. As readers would know, most fungi need this additional analysis for conclusive identification. As a reference book it would be nice to know if the author has lodged material as vouchers for the images to allow confirmation of some identifications. For example, none of the images on the Chanterelles page look typical to me. One specimen, which looks quite large and seems to be on wood, is not typical of a Chanterelle. I would also like to know more about the collection of *Podaxis pistillaris*, which is not found on termite mounds in Australia. However, *Podaxis beringamensis* is, and the published photo looks very similar to this species. It would be good to know if Australia and South Africa did share a termite mound fungi. Personally I have spent hours to no avail in Australia looking for *Termitomyces*, a termite mound genus common in Africa!

It is great to see images of many of the typical African macrofungi like *Termitomyces*, *Broomeia congregata* and *Lysurus corallocephalus*. There are also some examples of genera found in Australia such as *Lentinus* and *Amanita*. A number of species seem to be shared across these two localities including *Fomitopsis lilacinogilva*, *Polyporus arcularius*, *Leucocoprinus birnbaumii* and *Lentinus sajor-caju*. There are even some Fungimap target species (including *Aseroe rubra*, *Cyptotrama aspratium* and *Gymnopilus junonius*). Overall, the breadth of the macrofungi is excellent for a pocket guide.

While it is good to see a new book on the macrofungi of South Africa (the last available field guide was published in 1994 (Van der Westhuizen & Eicker, 1994)) my feeling is that this guide is perhaps not as well structured and researched as it could have been. Many of the fungi images are a bit soft or the fungi themselves are in poor condition. Nonetheless, it is great to see another field guide for South Africa. Hopefully an improved second edition will soon replace this edition. It would be a worthwhile purchase for those travelling to South Africa or those who want to see as many different fungi as possible.

**Reference**

Van der Westhuizen, G.C.A. and Eicker, A. (1994). *Mushrooms of Southern Africa*. Struik Publishers, Cape Town



MARIEKA GRYZENHOUT

There's no need to despair that there's too long to wait until the next Fungimap event!

Fungimap, in partnership with NRM South (Tasmania), will be running a **Fungi Festival in Hobart from Thursday April 26th to Monday April 30th, 2012.**

The proposed program is for the Thursday and Friday sessions focus on fungi conservation and management, and the Saturday and Sunday to consist of workshops and forays, similar to past Fungimap conferences. The Monday session is tentatively set aside for training in the use of the Atlas of Living Australia for fungi records data. The following weekend (5th-6th May), Fungimap, in conjunction with Cradle Coast NRM, will be running some more forays and workshops in the Tarkine.

Take note of the dates - more details to be announced soon!

## IS MARASMIELLUS AFFIXUS A LICHEN?

Heino Lepp

In *Fungimap Newsletter* 42 (p. 8) *Marasmiellus affixus* was considered to be a lichen. Fruit-bodies were described as growing in colonies on a mat 30 cm or larger consisting of a thin film of algal cells and mycelium (Catcheside *et al.*, 2011). In most lichens, the fungal component is an ascomycete. Of the 18,000 or so known lichens fewer than 100 are basidiolichens, that is those with a basidiomycete as the fungal component. Recently I needed to find out about the world's basidiolichens and I was surprised to find no proof of the lichenised status of *Marasmiellus affixus*. Kantvilas & May (1995) wrote that the 'lichenisation of this species was first noted by Singer (1970)' who stated '*Marasmiellus affixus* ... is lichenised and grows in association with a crustaceous lichen (*sic*) on the surface of *Eucalyptus* saplings in Australia' - but that is all. There is no reference to any research into a fungal-algal connection. A little later Singer (1973) gave more details and it's worth repeating his crucial comments because, although based on a single collection from New South Wales, they comprise the only detailed description of *Marasmiellus affixus*:

'I have received material of a crustaceous organism consisting of *Coccomyxa* [a genus of algae] imbedded in basidiomycetous hyphae, and these fruiting in the form of a pleurotoid *Marasmiellus* ... which was identified as *Marasmiellus affixus* ... by me. It was collected ... between Major's Creek and Araluen, ... on saplings of *Eucalyptus robertsonii*.

This *Marasmiellus* ... has a poor Rameales-structure [branched hyphae] in the surface layers of the pileus, and ellipsoid spores (5–)6.5–7.6 x 3.8–4.5  $\mu$ , inamyloid; the hyphae with walls 0.2–0.5  $\mu$  thick, with clamp connections. ...

The carpophores are gregarious and found exclusively where there is a covering of algae. The mycelial mat interweaving with the *Coccomyxa* is in places so scattered that in rather large accumulations of algal cells only a few mycelial threads can be seen whereas in others the mycelium is so thick that the algal cells are completely covered by it and the crustaceous base on top of the eucalyptus (*sic*) bark appears white instead of green. The crust is of indistinct morphology and therefore the association of alga and fungus appears to be less strict and obligatory than in *Botrydina* and *Coriscium* .... On the other hand some cells of the alga are occasionally found in the trama of the fruiting body.'

Ellipses indicate places where irrelevant collection details or taxonomic observations have been removed, but all Singer's descriptive information is retained. Explanatory information has been added in square brackets.

In the past *Botrydina* and *Coriscium* were accepted as proper generic names, occasionally to include species now placed in the genus *Lichenomphalia*. However, for good technical reasons that I'll leave unexplained, *Botrydina* and *Coriscium* have been rejected as formal generic names

but are used informally to denote two basidiolichen thallus types. *Botrydina*-type thalli are granular, the *Coriscium*-type are flake-like or scale-like and each granule or scale contains both photobiont cells and hyphae. Both types are found in the genus *Lichenomphalia*, but are not confined to that genus. There are basidiolichen thallus types other than those two. For example some *Dictyonema* species have thin but robust shelf-like thalli, each composed of a dense matrix of hyphally-sheathed photobiont filaments.

Oberwinkler (2001) gives a good summary of basidiolichen structure with diagrams that show the details of the fungus-photobiont connections in various species. *Marasmiellus affixus* is mentioned neither by Oberwinkler (2001) nor in the detailed molecular analyses of basidiolichens by Lawrey *et al* (2009). Singer published no observations about the nature of any hyphal-algal connection in *M. affixus* and, in essence, assumed lichenisation based on the proximity of hyphae and algal cells. Perhaps the alga simply creates an ideal microhabitat for the fungus but with no lichenisation involved. The occurrence of some algal cells in the trama of the fruit-body is not significant, since foreign bodies occasionally appear in fungal fruit-bodies.

So, what is the answer to the question posed in the title of this article? I seem to deal harshly with Singer and you may think I deny lichenisation. By no means so. Singer's comments are illuminating and suggest lichenisation. However, it is important to be aware of the absence of proof and all that one can say for sure is: 'plausible but not proven'. It is clear that *Marasmiellus affixus* would offer several interesting research projects involving molecular, cultural and ultrastructural techniques.

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## FUNGIMAP - PRESIDENT'S REPORT

### Year ending 31 December 2010

At the end of 2010 there were 215 paid up members (119 full, 70 concessional and 26 associate), which was an increase over the 174 members for 2009. Membership has thus returned to the level in 2008 (212 members), which is a pleasing result, because membership increases are usually greater in the Conference year (which was 2009). Fungimap has members from all states and the Northern Territory, as well as some overseas members.

In June, Fungimap farewelled Lee Speedy after two years as the Fungimap Coordinator. During the first half of the year Lee dealt with a wide variety of tasks including book sales and ordering and support for office volunteers. After Lee's departure, the position was temporarily placed on hold for the remainder of the year. This resulted in significant cost savings, but also an increased workload for volunteers in the Fungimap office and for the committee. A break in the continuity of the Fungimap Coordinator position has occurred on several occasions in the past. Whilst there are financial advantages, it is not a sustainable practice in the long term, because of the increased load on volunteers and the disruption to both day to day operations and to forward planning for activities such as conferences.

The Fungimap VI Conference will be held in Denmark in south-west Western Australia in July 2011, jointly organised with the Western Australian Field Naturalists Club. The Conference Organising Subcommittee comprises Sapphire McMullan-Fisher (chair), Joe Froudust, Roz Hart, Jolanda Keeble, Dawn Pedro, Lee Speedy (until June) and Katie Syme. The Subcommittee met regularly and preparations for the Conference are well underway, including confirmation of the venue as the Denmark College of Agriculture.

In the year in between Conferences Fungimap has traditionally held a low-key 'get-together' of the committee, to allow a face-to-face committee meeting (given that committee members are spread around Australia). In past years, this get together has been tied to expeditions and/or to relatively small scale talks and workshops. In 2010 the get together turned into a major activity in north-eastern Tasmania: 'Hidden treasures: discovering the fungi of the Blue Tier'. Sarah Lloyd (Central North Field Naturalists) and Katie Syme did much of the organising of the Blue Tier activities and financial support was provided by NRM North. The event was based in the town of Weldborough and included forays and workshops as well as an expedition to collect and document the fungi of the Blue Tier. More than 50 people attended the talks and workshops, and activities were also organised for 40 primary and secondary school students from the nearby town of St Helens. The Blue Tier event turned out to be quite similar in scope and format to the full-blown Conference that we hold every two years, the major difference being that attendees were responsible for their own food and accommodation and the workshops were at a more general level. The success of the Blue Tier activities opens up the possibility of more events, given suitable financial and logistical support at the local level.

Five new target species were 'launched' in 2010 (*Entoloma viridomarginatum*, *Filoboletus manipularis*, *Marasmiellus affixus*, *Podaxis beringamensis* and *Porodisculus pendulus*). Fungimap records continue to be submitted, with a total of 35,898 records logged in the Fungimap database at the end of 2010. These include the target species themselves but also records of non-target species, particularly from foray lists compiled by fungi interest groups, such as the Field Naturalists Club of Victoria Fungi Group. Some 4,507 records were added in 2010, which is an increase over the 2009 total of 3,215. However, there is a discrepancy between the figure at the end of 2009 as provided in the last annual report (32,814) and that used to calculate the 2010 figures (31,391). This highlights deficiencies in the reports used in the database and in the underlying programming. All original data is held on file in the Fungimap office and the overall quality of the data in the database is reasonably high. Nevertheless, considerable repairs and updates to the database which holds the Fungimap records are required. One example is making sure that each field is used only for the specific data it is intended (using a controlled set of options).

The Fungimap bookshop provides a service to members and is also an important source of funds, contributing about half of the income of the organisation in years when there is no conference. The Fungimap publication *Fungi Down Under* continues to sell steadily, with wholesale sales making up more than half the income from sales of the book. Of books and other items sold, around 80% of the income comes from *Fungi Down Under*.

Three issues of *Fungimap Newsletter* were prepared in 2010: issue 40 appeared in April (16 pp) and issue 41 in October (20 pp and two page colour insert), while the final issue for the year (42) was not distributed until January 2011 (16 pp and four page colour insert). A change introduced this year was the move from a single colour issue per year to use of colour pages in several issues. Sarah Lloyd very kindly laid out and printed the

colour pages and this significant support from Sarah and the Central North Field Naturalists is much appreciated. The colour section in issue 41 showcased interesting and beautiful fungi from Blue Tier and the colour pages in issue 42 highlighted the new target species.

The *Newsletter* continued as a venue for reports from fungi groups around Australia and also included a mix of short reports on interesting fungi or locations for foraging and articles about particular groups of fungi. The comprehensive calendar of fungi forays and other activities around Australia was printed only in the first issue for the year, and thereafter was kept up-to-date on the Fungimap website. The *Newsletter* was edited by Pam Catcheside and Tom May. Pam carried out much of the organisation behind the *Newsletter*, especially soliciting articles and information for the calendar of events and also doing the initial layout, which was finalised by Tom.

A *Fungimap Bulletin* was issued in April 2010, including articles on the choice of target species and phenology recording. The *Bulletin* was introduced as an irregular publication for issues and ideas beyond those dealt with in *Fungimap Newsletter*, and was made available through the Fungimap website.

Two committee meetings were held during the year, in March and May. All committee members were present at each meeting, and the whole committee actively contributed to the running of Fungimap. Exchanges of emails were used to deal with issues arising between meetings, where formal motions were required.

Volunteers are a vital part of Fungimap, both through the submission of records and images, but also in the Fungimap office. John Carpenter, Wendy Cook, Geoff Lay and Graham Patterson continued to provide support in the office, such as dealing with book orders, membership renewals, enquiries about the identity of fungi and logging and databasing records (especially over the period when there was no Fungimap Coordinator). Royal Botanic Gardens Melbourne continued to provide valuable assistance in hosting the Fungimap office and employed the Coordinator on behalf of Fungimap.

For 2010 there was a surplus of \$6,302. This represents a significant turnaround from 2009, when there was a loss of around \$1500. A decrease of expenditure on the Fungimap Coordinator (a combination of reducing to one day a week and the gap in the position) was the major contributing factor to the surplus, but there was also additional income from the Blue Tier activities as well as from the increase in donations and in membership numbers. The Austral Fungi Fund is the special fund that receives tax deductible donations under the Deductible Gift Recipient status granted to Fungimap. In 2010 donations to the Austral Fungi Fund totalled \$2,156, which is a significant increase over the 2009 donations of \$266. There was no expenditure from the Fund.

In 2011, the aim will be to build on the foundation of a financially successful Fungimap VI Conference to support continual employment of a part-time Fungimap Coordinator (2 days per week) and to seek additional funding for special projects related to the records database and publications.

**Tom May, President**

**Fungimap Inc No A 0047228L**  
**Statement of Financial Performance: January-December 2010**

**INCOME**

Memberships		\$5,729
Booksales:		
"Fungi Down Under" -Retail	\$3,515	
"Fungi Down Under" -Wholesale	\$4,074	
Books not "Fungi Down Under"	\$1,483	\$9,072
Fungimap CD-ROM		\$205
Sales of other stock		\$405
Handling and Postage: payment received		\$618
Bank Interest Earned		\$241
Donations		\$2,156
Donations - non DGR		\$100
Miscellaneous Income		\$436
<b>Total Income</b>		<b>\$18,962</b>

**EXPENDITURE**

Cost of Stock sold:		\$1,962
Replenish Inventory: Books	\$801	
Other	\$254	\$1,055
Book Stock Revaluation		\$236
Printing Newsletters and Brochure:		\$401
Insurances: Fungimap Volunteers		\$807
Administration Expense:		
Coordinator Salary and on-costs	\$6,606	
Travel	-	
Teleconferencing	\$389	
Office supplies and Stationery	\$136	
Subscription (Philanthropy Australia)	\$77	
Postage Paid & Couriers	\$7	
Bank Charges	\$158	
Merchant Card Charges	\$185	
Accounting fees	\$525	
Filing and Compliance Fees	\$41	\$8,124
<b>Total Expenditure</b>		<b>\$12,585</b>

**Blue Tier Expedition:**

<b>Income:</b>	\$6,510	
<b>Expense:</b>	\$5,221	
		<b>\$1,289</b>

<b>Fungimap Conference Denmark, 2011</b>		
<b>Deposit for Conference Venue</b>		<b>(\$1,364)</b>

<b>NET SURPLUS/(LOSS)</b>		<b>\$6,302</b>
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**Fungimap Inc No A 0047228L  
Statement of Financial Position**

<b><u>ASSETS</u></b>	<b>Dec 10</b>	<b>Dec 09</b>
<b>CASH AND AT BANK</b>		
Petty Cash: (held at the Fungimap office, Royal Botanic Gardens Melbourne)	\$4	\$48
Cash at Bank:		
Bendigo Bank Account 633-000 125124321	\$15,960	\$16,188
Austral Fungal Fund:	\$3,587	\$1,431
Investment Account (Bendigo Bank)	\$8,240	
GST Credits		\$331
Royal Botanic Gardens Melbourne, Fungimap Account (GST inclusive)		
<b>TOTAL CASH AND AT BANK:</b>	<b><u>\$27,791</u></b>	<b><u>\$17,998</u></b>
<b>OTHER ASSETS:</b>		
Stock of Books held at RBG: (Inventory taken 5 January)	\$8,269	\$10,396
Debtors and undeposited payments	\$348	\$377
<b>TOTAL ASSETS:</b>	<b><u>\$36,408</u></b>	<b><u>\$28,771</u></b>
<b><u>LIABILITIES</u></b>		
Creditors	\$2,114	\$1,122
GST Liability	\$324	
<b><u>NET ASSETS:</u></b>	<b><u>\$33,970</u></b>	<b><u>\$27,649</u></b>
<b><u>FUNGIMAP INC EQUITY:</u></b>		
B/f 31 December	\$27,668	\$28,913
Surplus/(Loss) from 'Financial Performance'	\$6,302	\$272 (\$1536)
<b><u>TOTAL EQUITY</u></b>	<b><u>\$33,970</u></b>	<b><u>\$27,649</u></b>

Fungimap Inc No. A 0047228L

Report of the Committee of Management

Your committee is pleased to submit the financial <sup>17</sup>accounts of Fungimap Inc for the calendar year January to December 2010.

The names of the committee members in office at the date of this report are:  
J Carpenter, P Catcheside, P George, T May, S McMullan-Fisher and K Syme  
The Public Officer of Fungimap Inc is Mr J Carpenter.

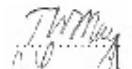
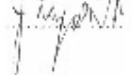
The principal activities and objects of Fungimap Inc. are to promote and support the study and conservation of Australian macrofungi.

The net surplus for the twelve month period is \$6,302. No provision for income tax is required, as Fungimap Inc has been self-assessed as income-tax exempt.

No office holder has received or become entitled to receive, during or since the end of the reporting year, a benefit because of a contract made by Fungimap Inc with the office holder or any entity with which the office holder has a substantial interest.

During or since the end of the reporting year, Fungimap Inc has not entered into any mortgage or other arrangements affecting any of the property of the association. Fungimap Inc has not created any trusts, and is not a trustee of any trust.

Signed on T. May in accordance with a resolution of the Committee of Management.

Director  T May (President)  
Director  J Carpenter (Treasurer)

Statement by the Committee of Management

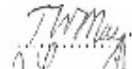
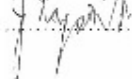
The office holders declare:

(1) that the following financial statements and notes give a true and fair view of the financial position of Fungimap Inc for the 12 month period 1 January 2010 to 31 December 2010, and of the financial performance of Fungimap Inc for that period;

(2) that at the date of this statement, there are reasonable grounds to believe that the association will be able to pay its debts as and when they fall due;

(3) that in the intervening period between 31 December 2010 and the date of this declaration, there have been no material changes to the affairs of the association.

Signed on T. May in accordance with a resolution of the Committee of Management.

Director  T May (President)  
Director  J Carpenter (Treasurer)

**ACKNOWLEDGEMENTS**

**RECORDS**

**AUSTRALIA (by email)**

Philip Bridle	1
Steve Burrows	1
Kath Matthews	2
Katherine Tuckett	1

**NSW**

Martin & Frances Butterfield	6
Judith Carrick	5
Bill Crozier	1
John D	1
Ben Reid	1
Sydney Fungal Studies Group	15
Teresa Van Der Heul	51
Sarah Wilks	1

**QLD**

Rod Hobson	2
Sapphire McMullan-Fisher	21

**SA**

Adelaide Fungal Studies Grp	58
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Kath Alcock	1
David Catcheside	2
Julia Haska	22

**TAS**

Carolyn Hall-Jones	48
Patricia Harrison	72
Nell Hilliard	8
Sarah Lloyd	189
Lori Puster	1
Roy Skabo	21

**VIC**

Robert Bender	20
Wendy Cook	17
Don Greaves	1
Cecily Falkingham	49
Field Naturalists Club of Vic	254
Geelong Field Naturalists Club Inc	2
Paul George	2
Sally Green	28

Cath Greenop	9
Jean Lightfoot	25
Ivan Margitta	57
Malcolm McKinty	10
Catherine Nield	1
Win Pietsch	9
Rowan	2
Sale and District Field Naturalists Club	11
Nigel Sinnott	3
Faye Vyner	59

**WA**

Neale Bougher	1
Michael Gregson	1
Lee Hollingsworth	1
Jan Lewis	1
James Pillsbury	1

**VOLUNTEERS**

Thanks to Royal Botanic Gardens Melbourne and our volunteers: John Carpenter, Wendy Cook, Geoff Lay and Graham Patterson for providing office and administrative support

**FUNGIMAP**

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This *Fungimap Newsletter* was edited by Christina Hall, Pam Catcheside and Tom May.

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**FUNGIMAP**

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